



**IJVR** 

ISSN: 1728-1997 (Print) ISSN: 2252-0589 (Online)

Vol. 23

No. 2

Ser. No.79

2022

# IRANIAN JOURNAL OF VETERINARY RESEARCH



### **Original Article**

# Application of histochemical and immunohistochemical techniques for detection of lung tissue in cooked sausage

Sami, M.1\*; Kheirandish, R.2; Nasri, A.3 and Dabiri, Sh.4

<sup>1</sup>Department of Food Science and Technology, School of Nutrition and Food Science, Isfahan University of Medical Sciences, Isfahan, Iran; <sup>2</sup>Department of Pathobiology, School of Veterinary Medicine, Shahid Bahonar University, Kerman, Iran; <sup>3</sup>Department of Veterinary Biomedical Sciences, Western College of Veterinary Medicine, University of Saskatchewan, Saskatoon, Canada; <sup>4</sup>Department of Pathology, School of Medicine, Pathology and Stem Cell Research Center, Afzalipour Hospital, Kerman University of Medical Sciences, Kerman, Iran

\*Correspondence: M. Sami, Department of Food Science and Technology, School of Nutrition and Food Science, Isfahan University of Medical Sciences, Isfahan, Iran. E-mail: masoud\_sami@nutr.mui.ac.ir



10.22099/IJVR.2022.40333.5849

(Received 12 Apr 2021; revised version 5 Feb 2022; accepted 12 Feb 2022)

### **Abstract**

Background: Using unauthorized tissues in sausage is a common food adulteration in some parts of the world. Aims: This study was designed to compare the accuracy of histochemical and immunohistochemical techniques for the detection of lung tissue in cooked sausage samples. Methods: Samples with different levels of sheep lung tissues (1, 2.5, and 5%) and a control group were prepared and stained histochemically using H&E, Masson trichrome, and Periodic Acid-Schiff (PAS) stainings, and immunohistochemically using two different commercially-available antibodies of TTF1 Pan-cytokeratin. Results: The highest positive results of lung tissue detection were achieved in sausage samples stained with anti-TTF1 immunohistochemical staining method. Both anti-TTF1 and anti-pan-cytokeratin immunohistochemical techniques detected all contaminated sausage samples treated with 50 g/kg lung tissues. Anti-TTF1 staining method had the highest odds ratio (7.4), followed by anti-pan-cytokeratin method (6.0). Reversely, PAS staining method had the lowest odds ratio (0.21), followed by Masson trichrome method (1.7). Additionally, anti-TTF1 method had the highest (1.8-31.0) confidence intervale (95%), while PAS had the lowest (0.02-2.1). Totally, the odds ratio of lung tissue detected by immunohistochemical methods were higher than those detected by histochemical staining. Conclusion: This is the first report on the comparison of histochemical and immunohistochemical techniques for lung tissue detection in cooked sausage. Anti-TTF1 immunohistochemical staining proved to be the most useful technique for the detection of unauthorized lung tissue in cooked sausages.

Key words: Food adulteration, Immunohistochemistry, Lung tissue, Sausage

### Introduction

Sausage is an acceptable ready-to-eat food product among all age groups, worldwide. The consumption of sausage has increased, and therefore consumers need assurance regarding the quality and safety of sausage products. In spite of many efforts of producers to improve the quality of sausage, the public's view has been partly negative about these ready-to-eat products due to recent issues raised in food adulteration and recent negligence about the presence of contaminated central nervous system (CNS) with bovine spongiform encephalopathy (BSE) in meat products (Chen *et al.*, 2013; Pandey *et al.*, 2020).

Different types of undesirable and sometimes detested ingredients among customers are added to meat products because of their low price or technological properties in different processes of sausage production (Ahmed *et al.*, 2020). Strikingly, a worldwide increase in food prices especially for meat has caused some producers to use undesirable tissues such as the spleen,

liver, mammary gland, and lung in meat products (Dehghan Shahreza, 2016). Since animal offal can be contaminated with bacteria, viruses, foreign bodies, and parasites or even can be served as a pathway for various types of environmental toxicants to enter the human body, the use of these resources in meat products should be monitored more carefully. Given this point for lung tissue, some studies suggested that the significant uptake of cadmium (Cd) and manganese (Mn) in dairy farms can happen through the lung (Sunderman, 2001; Bressler et al., 2004; Roggeman et al., 2014; ShahbaziGahrouei and Keshtkar, 2016). Also, the animal lung can be a potential vehicle for food-borne bacteria including listeria (Kuan et al., 2013). Thus, awareness about animal offal including lung can play an important role in food quality and safety control, especially due to the European Union's strict regulations on the use of specified risk material (SRM) (2000/418/EC), their agreement for labeling all the permitted edible parts (2001/101/EC) and more careful inspection of the used offal in meat products (Sultan et al., 2004).

The importance of efficient techniques for evaluating the composition of comminuted meat products has increased (Koolmees and Bijker, 1985; Koolmees et al., 1986; Meret et al., 1998; Hajmeer et al., 2003), for example, immunochemical and immunohistochemical methods have been used to detect CNS as unauthorized tissue in commercial beef sausages or homemade meat products (Boon, 1990; Lücker et al., 1999; Schmidt et al., 1999; Lücker et al., 2000). Regarding this point, there have been some commercial kits for the detection of CNS tissue in meat products (Yeşilbağ and Kalkan, 2005). However, no kit has been designed for the detection of other tissues. Hence, the present research was conducted to the comparison of the performance of histochemical and immunohistochemical techniques for the detection of the lung tissue as adulteration in cooked sausage.

### **Materials and Methods**

### Reagents

The anti-thyroid transcription factor 1 (TTF1) and Pan-cytokeratin antibodies were obtained commercially from Dakocytomation Company, Denmark.

### Sausage production

The production procedure of a lyoner meat-type product was as follows: after mincing calf meat with a mincer machine with 3 mm plates of stainless steel (Pars Khazar Co., Iran), they were mixed with ice, salt, ascorbate and phosphate in a food maker (Moulinex Co., France) until the temperature reached 10°C. Then, Soya, flour, starch, and spices were added to the mixture and the final mixture was chopped until a suitable consistency was achieved at a temperature of 15°C. This mixture was considered as the control group (group A), then different amounts of minced lung tissue were added to the chopped product to form groups B, C, and D including 10, 25, and 50 g/kg lung tissue, respectively. They were mixed again, filled to the commercial artificial casing, and located in a water bath with a core temperature of 72°C for 2.5 h. Experimental sausage production was repeated 5 times.

### **Histochemical staining**

From each sausage, one block ( $1 \times 1 \times 1$  cm) was taken. A total of 20 blocks were sampled from 4 different groups. Samples were then fixed in a 10% neutral buffered formalin solution and processed according to the routine paraffin wax method. From each Paraffin block, 5 sections ( $5 \mu m$ ) were cut. Three sections were stained histologically by hematoxylin and eosin (H&E), Masson trichrome, and Periodic Acid-Schiff (PAS). Totally, 100 sections were prepared and all fields of each slide were carefully examined with magnification of  $\times 100$  and  $\times 400$  by a light microscope.

### **Immunohistochemical staining**

The tissue sections were stained immunohistochemically using an avidin-biotin peroxidase complex (ABC) method (Haines and Chelack, 1991). The sections were deparaffinized and rehydrated by sequential immersion of the slides in xylene, graded concentrations of ethanol and distilled water. Then slides were immersed in a citrate solution and heated in a microwave oven for 3 min until the solution started to boil. Endogenous tissue peroxidase activity was blocked by immersion of the slides in a solution of 0.5% H<sub>2</sub>O<sub>2</sub> in methanol for 30 min at room temperature. Sections were treated with phosphate-buffered saline supplemented with 4.0% normal goat serum to block non-specific background staining, then sections were incubated with monoclonal antibodies such as TTF1 and Pan-cytokeratin with a concentration of 1:100 in PBS for 60 min. After washing (3 times and 5 min for each time) with PBS/Tween, sections were incubated with a biotinylated goat anti-rabbit antibody for 30 min. They were washed with PBS/Tween, flooded with avidinbiotin complex peroxidase solution and again were washed 3 times with PBS/Tween. The peroxidase activity was visualized with 1 mg/ml DAB (3, 3'-Diamino-Benzidine) in PBS supplemented with H<sub>2</sub>O<sub>2</sub> (10 µL of 50% H<sub>2</sub>O<sub>2</sub> in 5 ml PBS) as a chromogen. After washing with PBS/Tween, sections were counterstained with Mayer's hematoxylin for 1 min. Finally, they were dehydrated, mounted and examined by light microscopy.

### Statistical analysis

The ratio of positive-diagnosed slides to the total slides of each staining was considered for further statistical analysis. Then, the logistic regression model was used to calculate odds ratios (OR, multiplicative), corresponding 95% confidence intervals (CI) and significant differences between histochemical techniques. These methods were considered reference methods and immunohistochemical methods were compared with them.

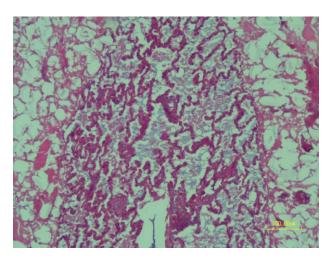
### **Results**

### **Analysis of staining methods**

The current survey was done to evaluate the accuracy of histochemical and immunohistochemical techniques to detect unauthorized lung tissue in sausage samples. Sausage samples with different amounts of lung tissue were stained with three staining methods.

Figure 1 signifies the findings of the H&E staining method for the detection of lung tissues. Alveolar-like structure with thin septa was seen in the H&E staining method which was considered positive for lung tissue. In H&E sections of the control group, some homogenous eosinophilic materials and skeletal fibers were observed, but in the experimental groups, thin-walled and alveolar-like structures as cleft-like features were identified as well as skeletal fibers with degenerative changes (Fig. 1). Figure 2 signifies the findings of the Masson trichrome staining method for the detection of lung tissue in studied sausage samples. More clear alveolar-like structure and hyaline cartilage were found in sausage samples in Masson trichrome staining method which was considered

positive for lung tissue. In Masson trichrome staining, alveolar ducts were stained red to purple, and connective tissue between alveolar septa and hyaline cartilage was stained greenish blue (Fig. 2). Figure 3 signifies the findings of the PAS staining method for the detection of lung tissue in studied sausage samples. Carbohydrate ingredients were stained as dim brownish which was considered positive for lung tissue. Figure 4 signifies the findings of the anti-TTF1 immunohistochemical staining method for the detection of lung tissue in the studied sausage samples. The nuclei of pneumocytes and Clara cells were found darker than normal tissues which were considered positive for lung tissue. Figure 5 signifies the findings of the anti-pan-cytokeratin immunohistochemical staining method for the detection of lung tissue in studied sausage samples. Positive immunoreactions of airway epithelial cells were considered positive for lung tissue. Figure 6 signifies the findings of the H&E staining of sausage samples to determine the presence of plant structure and striated muscles.



**Fig. 1:** Group C. This photomicrograph shows a section of lung tissue containing alveolar structures with thin walls, (H&E, scale bar: 100 μm)

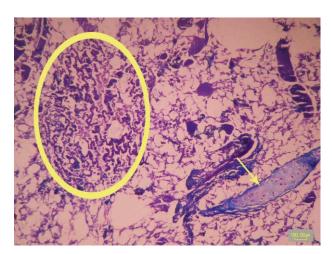


Fig. 2: Group D. Presence of Alveolar structures (circle) and hyaline cartilage (arrow), (Masson trichrome, scale bar: 100  $\mu$ m)

## Comparison of the accuracy of staining techniques

Table 1 signifies the results of the histochemical and

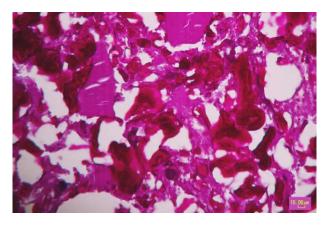
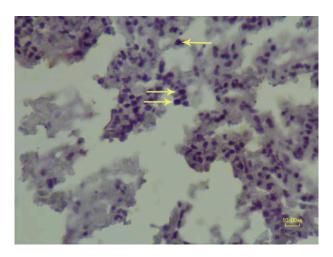


Fig. 3: Group C. Carbohydrate ingredients stained as dim brownish and disappear other tissues, (PAS, scale bar:  $10\,\mu m$ )



**Fig. 4:** Group B. The nuclei of pneumocytes and Clara cells are darker than normal ones (arrows), (Anti-TTF1 immunohistochemical staining, scale bar: 10 μm)

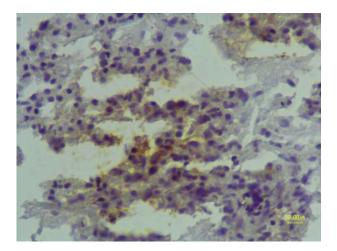
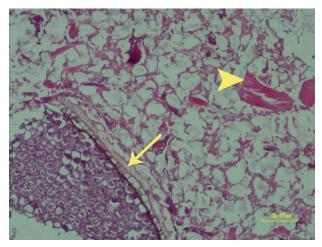


Fig. 5: Group B. Positive immunoreactions of airway epithelial cells related to lung tissue are seen, (Anti-pan-cytokeratin immunohistochemical staining, scale bar:  $10~\mu m$ )



**Fig. 6:** Group C. Presence of plant structure (arrow) and striated muscles (arrowhead) in the sausage section, (H&E, scale bar: 100 µm)

immunohistochemical techniques in the detection of lung tissue. Among all histochemical tests, Masson trichrome staining had the highest accuracy in the detection of lung tissue in cooked sausage samples. Among two studied methods, TTF1 technique had higher accuracy only in sausage samples treated with 10 g/kg lung tissue. Additionally, TTF1 and Pan-cytokeratin immunohistochemical techniques both detected all contaminated sausage samples treated with 50 g/kg lung tissues. However, Masson trichrome staining cauld detect only 3 contaminated age samples treated with 50 g/kg lung tissues. Table 1 shows the number of positive results in both histochemical and immunohistochemical methods. In Masson trichrome staining, the rate of the lung tissue diagnosed positively, was higher than positive results of another histochemical staining, except for the 50 g/kg added lung tissue in which approximately 60% of the slides stained with H&E and Masson trichrome, were detected positively.

Table 2 signifies the accuracy of different staining methods for detection of lung tissues in studied sausage samples. H&E staining method was considered as a reference. Comparison of the odds ratio of studied techniques revealed that TTF1 staining method had the highest odds ratio (7.4), followed by the Pan-cytokeratin method (6.0). Reversely, PAS staining method had the lowest odds ratio (0.21), followed by Masson trichrome method (1.7). Comparison of the confidence intervale (95%) disclosed that TTF1 method had the highest (1.8-31.0) confidence intervale, while PAS had the lowest (0.02-2.1). In confirmation of the mentioned results in

Table 1, the odds ratio of lung tissue detection for Masson trichrome staining was more than other histochemical methods, but this difference was not significant (P>0.05, Table 2).

**Table 2:** Comparison of the accuracy of different methods for the detection of lung tissue in sausage samples

Detection methods	Odds ratio	CI (95%)	P-value
H&E	1 (Reference)	-	-
Masson trichrome	1.7	0.4-7.3	0.5
PAS	0.21	0.02-2.1	0.2
Anti-TTF1	7.4	1.8-31.0	0.006
Anti-pan-cytokeratin	6.0	1.5-24.6	0.013
Histochemical	1 (Reference)	-	-
Immunohistochemical	7.4	3.0-18.5	0.001

### **Discussion**

Although numerous histochemical studies have been applied to determine whether unauthorized tissues could be detected in the meat products, our results were not in agreement with those of previous studies which reported that histochemical staining was useful to confirm the presence of unauthorized animal tissues in food products (Lazzaro et al., 1991; Prayson et al., 2008a, b; Sadeghinezhad et al., 2015). In the present study, lung structures were seen relatively distinctive using three histochemical staining methods, but in Periodic Acid-Schiff staining, the lung tissues were not detected due to the high contrast of added carbohydrates in sausage which created a dim brownish pattern. Generally, identification of lung tissue by H&E, Masson trichrome and Periodic Acid-Schiff staining in cooked samples was not confirmed. Thus, standard light microscopy was not adequate to detect small autolytic fragments of lung tissue and did not allow discrimination between fragments of mixed materials (a normal component of meat).

According to the Iranian National standard regulations, the use of undesirable organs of slaughtered animals, including the skin, visceral organs, hyaline cartilage, bone, and fat instead of meat in meat products is considered adulteration (Moghtaderi *et al.*, 2019). Numerous researches were addressed for disclosure of unauthorized tissues in meat products (Cetin *et al.*, 2016; Moghtaderi *et al.*, 2019). Investigations on heated meat products demonstrated the presence of adipose tissue, plant material, peripheral nerves, blood vessels, cartilage, bone, gizzard, lymph node, lung tissue, gland tissue, udder tissue, ovary, and cartilage (Latorre *et al.*, 2015).

Table 1: Results of the histochemical and immunohistochemical methods for detection of lung tissue

Lung tissue addition (g/kg)	Histochemical methods (positive results/total tested samples)		Immunohistochemical methods (positive results)		
	H&E	Masson trichrome	PAS	TTF1	Pan-cytokeratin
0	0/5	0/5	0/5	0/5	0/5
10	0/5	1/5	0/5	3/5	2/5
25	1/5	2/5	0/5	5/5	5/5
50	3/5	3/5	1/5	5/5	5/5

Immunohistochemical methods are based on the presence of targeting reactive antigens in sections. This technique is reliable and quite sensitive, and therefore it can be used in different aspects of food safety including the detection of food components especially harmful ingredients, the adulteration of foodstuffs and quantification of food ingredients (Kalčáková *et al.*, 2021).

In our study, two characteristic protein markers including TTF1 and Pan-cytokeratin were used to detect the lung in cooked samples. Nowadays TTF1 has been considered as a reliable marker to distinguish lung tissue. This antibody is a 38-kDa homeodomain containing DNA-binding protein of the Nkx-2 gene family. It is expressed in epithelial cells in the lung-like type II pneumocytes and Clara cells. The role of TTF-1 in lung involves the regulation of gene expression of surfactant and Clara cell secretory protein (Lazzaro et al., 1991; Jagirdar, 2008). Hence in the present research, immunostaining by TTF1 antibody detected TTF1 protein in the nucleus of pneumocytes and Clara cells in alveoli and bronchiole, respectively. Another protein, cytokeratin, is an intermediate filament structural protein, which is found in the cytoskeleton of various epithelial tissues such as airways in lungs (Ring et al., 2009). Due to different amounts of added lung tissue, different degrees of staining intensity could be observed. Whereas samples containing 25 g/kg of lung tissue had strong staining, the tissue samples with 10 g/kg lung showed less intensive staining. Cooked sausages without lung tissue revealed no immunoreactions. Generally, positive results obtained by immunostaining by TTF1 seemed to be more accurate than results obtained by Pancytokeratin, because the odd ratios of immunohistochemical staining by TTF1 was higher immunohistochemical staining by Pan-cytokeratin. In this way, the immunohistochemical techniques were more reliable in the detection of lung tissue compared to histochemical methods. In another survey (Moghtaderi et al., 2019) sausage sections were stained using H&E, Masson's trichrome, Periodic Acid-Schiff/Alcian blue, and Verhoeffe/Van Gieson to detect unauthorized tissues. A wide range of unauthorized tissues was detected, such as dense connective tissue (6.66%), cartilage (28.30%), bone (8.30%), skin (51.60%), smooth muscle (1.66%), and blood vessels (11.66%). They introduced the Masson's trichrome staining as a practical technique for routine assessment of authenticity and quality of sausage to protect the consumers from adulteration. Sadeghi et al. (2011) examined 720 sausage samples and found lung unauthorized tissues in about 4% of examined samples using histology. Sadeghinezhad et al. (2016) verified the efficacy of MT blue staining for detecting animal and herbal additive tissue in minced meat as well as common H&E staining. Pospiech et al. (2009) showed that some special stainings such as the PAS/Calleja staining which targets polysaccharides can indicate soybean flour. Gürbüz et al. (2020) reported the higher sensitivity and specificity of immunohistochemical methods (AE1/AE3 cytokeratin antibody) compared to histological examination in detection of adulteration in fermented sausage samples.

In agreement with our observation for immunohistochemical detection of lung tissue in cooked samples, the result of some studies showed different staining intensities by immunohistochemical detection of CNS in cooked sausages with different levels of brain tissue (Wenisch *et al.*, 1999; Tersteeg *et al.*, 2002). Also, the presence of spinal cord in Advanced Meat Recovery Systems (AMRS) by immunohistochemical and histochemical staining and polarization microscopy were proved (Kelley *et al.*, 2000).

The amount of immunoreaction in food samples is completely based on the state of food (raw, pasteurized and sterilized) and antibody resources even with the same amount of added target tissues (Tersteeg *et al.*, 2002). In this regard, one of the most important results of the present study was the ability of antibodies to detect lung tissue in final heated products. This result can be attributed to the heating process in which new epitopes can be generated in samples and used as a new target for new and improved antibodies (Tersteeg *et al.*, 2002).

The present study was limited to the lack of the evaluation of the role of molecular techniques for comparison with histological and immunohistochemical methods in the detection of fraud, as well as the lack of evaluation of other commonly used antibodies in the detection of fraud through immunohistochemical tests. Also, due to the absence of raw sausages in Iran, another limitation of this study is the lack of comparison of fraud detection methods on raw sausage samples.

In conclusion, the immunohistochemical procedure has many benefits including less time, less difficulty, and immunoreaction specificity. Beyond histochemical staining can be considered an inefficient method due to the high degree of tissue destruction (Pospiech et al., 2011). In the present study, all of the positive samples were detected by immunohistochemical technique compared to histochemical staining, and thus the accuracy of immunohistochemistry makes it very distinctive from conventional and daily pathological staining. Because adding lung tissue to meat products can create a threat for transmission of many diseases from infected animals to humans, national agencies need to apply more strict controls and surveillance and improve their hazard analysis and critical control points.

### Acknowledgement

This research was financially supported by the Research Council of Shahid Bahonar University of Kerman, Iran.

### **Conflict of interest**

The authors declare that they have no conflict of interest.

### References

- Ahmed, AM; Ismail, TH; Abouelmaatti, RR; Gaafar, RE and Elfeil, WM (2020). Detection of commercial fraud in processed meat products using rapid techniques. Am. J. Biochem. Biotechnol., 16: 244-251.
- **Boon, A** (1990). Histomorphometry and immunohistochemistry of beef sausages. J. Clin. Pathol., 43: 435.
- Bressler, JP; Olivi, L; Cheong, JH; Kim, Y and Bannona, D (2004). Divalent metal transporter 1 in lead and cadmium transport. Ann. NY. Acad. Sci., 1012: 142-152.
- Cetin, O; Bingol, EB; Civan, E; Turgay, SI and Ergun, O (2016). Identification of animal species and foreign tissues in ready-to-sell fresh processed meat products. Acta Aliment., 45: 198-205.
- Chen, CC; Wang, YH and Wu, KY (2013). Consumption of bovine spongiform encephalopathy (BSE) contaminated beef and the risk of variant Creutzfeldt-Jakob disease. Risk Anal., 33: 1958-1968.
- **Dehghan Shahreza, F** (2016). From oxidative stress to endothelial cell dysfunction. J. Prev. Epidemiol., 1: e04.
- **European Commission** (2000). Commission Decision 2000/418/EC regulating the use of material presenting risks as regards transmissible spongiform encephalopathies and amending Decision 94/474/EC. Off. J. Eur. Comm., L158, 76-82.
- **European Commission** (2001). Commission Directive 2001/101/EC of 26 November 2001 amending Directive 2000/13/EC of the European Parliament and of the Council on the approximation of the laws of the Member States relating to the labelling, presentation and advertising of foodstuffs. Off. J. Eur. Comm., L310, 19-21.
- Gürbüz, S; Ekebaş, G; Bayram, LÇ and Kaplan, YZ (2020). Quality determination of traditional fermented sausages by histological and immunohistochemical analyses. Akademik Gıda. 18: 288-295.
- Haines, DM and Chelack, BJ (1991). Technical considerations for developing enzyme immunohistochemical staining procedures on formalin-fixed paraffinembedded tissues for diagnostic pathology. J. Vet. Diagn. Invest., 3: 101-112.
- **Hajmeer, M; Cliver, DO and Provost, R** (2003). Spinal cord tissue detection in comminuted beef: comparison of two immunological methods. Meat Sci., 65: 757-763.
- **Jagirdar, J** (2008). Application of immunohistochemistry to the diagnosis of primary and metastatic carcinoma to the lung. Arch. Pathol. Lab. Med., 132: 384-396.
- Kalčáková, L; Pospiech, M; Tremlová, B; Javůrková, Z and Chernukha, I (2021). Development of immunohistochemical methods for casein detection in meat products. Foods. 10: 1-14. https://dx.doi.org/10.3390/foods 10010028.
- Kelley, LC; Hafner, S; McCaskey, PC; Sutton, MT and Langheinrich, KA (2000). An evaluation of methods for the detection of spinal cord in product derived from advanced meat recovery systems. J. Food Prot., 63: 1107-1112
- Koolmees, P and Bijker, P (1985). Histometric and chemical methods for determining collagen in meats. Vet. Quart., 7: 84-90
- Koolmees, P; Bijker, P; Van Logtestijn, J and Tuinstra-Melgers, J (1986). Histometrical and chemical analysis of mechanically deboned pork, poultry and veal. J. Anim. Sci., 63: 1830-1837.
- Kuan, CH; Wong, WC; Pui, CF; Mahyudin, NA; Tang, JYH; Nishibuchi, M and Radu, S (2013). Prevalence and

- quantification of Listeria monocytogenes in beef offal at retail level in Selangor, Malaysia. Braz. J. Microbiol., 44: 1169-1172.
- Latorre, R; Sadeghinezha, J; Hajimohammadi, B; Izadi, F and Sheibani, MT (2015). Application of morphological method for detection of unauthorized tissues in processed meat products. J. Food Qual. Hazard Control. 2: 71-74.
- Lazzaro, D; Price, M; de Felice, M and Di Lauro, R (1991).
  The transcription factor TTF-1 is expressed at the onset of thyroid and lung morphogenesis and in restricted regions of the foetal brain. Development. 113: 1093-1104.
- Lücker, E; Eigenbrodt, E; Wenisch, S; Failing, K; Leiser, R and Bülte, M (1999). Development of an integrated procedure for the detection of central nervous tissue in meat products using cholesterol and neuron-specific enolase as markers. J. Food. Prot., 62: 268-276.
- Lücker, E; Eigenbrodt, E; Wenisch, S; Leiser, R and Bülte, M (2000). Identification of central nervous system tissue in retail meat products. J. Food Prot., 63: 258-263.
- Meret, V; Guizard, C; Leduc, V; Nguyen, T; Da-Riz, V and Demeulemester, C (1998). Identification of animal and plant proteins in food ingredients and meat products. In: Proceedings of 44th International Congress of Meat Science and Technology. 30 August-4 September 1998, Barcelona, Spain. PP: 598-599.
- Moghtaderi, A; Raji, A; Khanzadi, S and Nabipour, A (2019). Application of histological method for detection of unauthorized tissues in meat sausage. Vet. Res. Forum., 10: 357-360.
- Pandey, P; Vidyarthi, SK; Vaddella, V; Venkitasamy, C; Pitesky, M; Weimer, B and Pires, AF (2020). Improving biosecurity procedures to minimize the risk of spreading pathogenic infections agents after carcass recycling. Front. Microbiol., 11: 1-13.
- Pospiech, M; Tremlova, B; Renčová, E; Lukášková, ZŘ and Pokorná, J (2011). Comparison of the results of the ELISA, histochemical, and immunohistochemical detection of soya proteins in meat products. Czech. J. Food. Sci., 29: 471-479.
- Pospiech, M; Tremlova, B; Renčová, E and Randulová, Z (2009). Immunohistochemical detection of soya protein-optimization and verification of the method. Czech. J. Food. Sci., 27: 11-19.
- Prayson, B; McMahon, J and Prayson, R (2008a). Applying morphologic techniques to evaluate hotdogs: what is in the hotdogs we eat? Ann. Diagn. Pathol., 12: 98-102.
- **Prayson, B; McMahon, J and Prayson, R** (2008b). Fast food hamburgers: what are we really eating? Ann. Diagn. Pathol., 12: 406-409.
- Ring, BZ; Seitz, RS; Beck, RA; Shasteen, WJ; Soltermann, A; Arbogast, S; Robert, F; Schreeder, MT and Ross, DT (2009). A novel five-antibody immunohistochemical test for subclassification of lung carcinoma. Modern Pathol., 22: 1032-1043.
- Roggeman, S; de Boeck, G; De Cock, H; Blust, R and Bervoets, L (2014). Accumulation and detoxification of metals and arsenic in tissues of cattle (*Bos taurus*), and the risks for human consumption. Sci. Total Environ., 466: 175-184.
- Sadeghi, E; Khazaei, M; Almasi, A; Shariatifar, N; Bohlouli Oskoii, S and Tahvilian, R (2011). Recognition of illegal tissues in the meat products from Kermanshah Supply Centers during the years 2009-2010. Horizon. Med. Sci., 17: 55-59.
- Sadeghinezhad, J; Hajimohammadi, B; Izadi, F; Yarmahmoudi, F and Latorre, R (2015). Evaluation of the morphologic method for the detection of animal and

- herbal content in minced meat. Czech. J. Food. Sci., 33: 564-569.
- Schmidt, G; Hossner, K; Yemm, R; Gould, D and O'Callaghan, J (1999). An enzyme-linked immunosorbent assay for glial fibrillary acidic protein as an indicator of the presence of brain or spinal cord in meat. J. Food. Prot., 62: 394-397.
- ShahbaziGahrouei, D and Keshtkar, M (2016). Magnetic nanoparticles and cancer treatment. Immunopathol. Persa., 2: e03.
- Sultan, KR; Tersteeg, MH; Koolmees, PA; de Baaij, JA; Bergwerff, AA and Haagsman, HP (2004). Western blot detection of brain material in heated meat products using myelin basic protein and neuron-specific enolase as

- biomarkers. Analyt. Chim. Acta. 520: 183-192.
- **Sunderman, FW** (2001). Nasal toxicity, carcinogenicity, and olfactory uptake of metals. Ann. Clin. Lab. Sci., 31: 3-24.
- **Tersteeg, M; Koolmees, P and Van Knapen, F** (2002). Immunohistochemical detection of brain tissue in heated meat products. Meat Sci., 61: 67-72.
- Wenisch, S; Lücker, E; Eigenbrodt, E; Leiser, R and Bülte, M (1999). Detection of central nervous tissue in meat products-An immunohistochemical approach. Nutr. Res., 19: 1165-1172.
- Yeşilbağ, K and Kalkan, A (2005). Detection of central nervous system tissues as BSE specified risk material in meat products in Turkey. Food Control. 16: 11-13.